

SELECTIVITY OF SEROTONERGIC DRUGS FOR MULTIPLE BRAIN SEROTONIN RECEPTORS

ROLE OF [³H]-4-BROMO-2,5-DIMETHOXYPHENYLISOPROPYLAMINE ([³H]DOB), A 5-HT₂ AGONIST RADIOLIGAND

MILT TITELER,*† ROBERT A. LYON,* KATHARINE H. DAVIS* and RICHARD A. GLENNON‡

*Department of Pharmacology and Toxicology, Albany Medical College, Albany, NY 12208; and

‡Department of Medical Chemistry, Virginia Commonwealth University, Richmond, VA 23298, U.S.A.

(Received 11 December 1986; accepted 3 March 1987)

Abstract—The affinities of putative serotonin receptor agonists and antagonists for 5-HT_{1A}, 5-HT_{1B}, 5-HT_{1C}, and 5-HT₂ receptors were assayed using radioligand binding assays. The 5-HT₁ sites were labeled with the agonist radioligands [³H]-8-hydroxy-2-(di-*n*-propylamino)-tetralin [³H]-8-OH-DPAT, [³H]-5-HT, and [³H]mesulergine. The 5-HT₂ receptor was labeled with the antagonist radioligand [³H]ketanserin or the agonist radioligand [³H]-4-bromo-2,5-dimethoxyphenylisopropylamine ([³H]DOB). The apparent 5-HT₁ receptor selectivity of agonist compounds was found to be 50- to 100-fold higher when the 5-HT₂ receptor affinity was determined using the antagonist radioligand [³H]ketanserin than when the agonist radioligand [³H]DOB was used. Quipazine, a putative specific 5-HT₂ agonist, appeared to be only 3-fold more potent at 5-HT₂ than at 5-HT_{1A} receptors when [³H]ketanserin was used as the 5-HT₂ radioligand. When [³H]DOB was used as the 5-HT₂ radioligand, quipazine was determined to be 100-fold more potent at 5-HT₂ receptors than at 5-HT_{1A} receptors. 1-(3-trifluoromethylphenyl)piperazine (TFMPP), a putative specific 5-HT_{1B} receptor agonist was apparently 10-fold more potent at 5-HT_{1B} receptors than at 5-HT₂ receptors when [³H]ketanserin was used as the 5-HT₂ radioligand. When [³H]DOB was used as the 5-HT₂ radioligand, TFMPP was found to be equipotent at 5-HT_{1B} and 5-HT₂ receptors. Using the 5-HT₂ antagonist radioligand [³H]ketanserin, a similar pattern of underestimating 5-HT₂ receptor selectivity and/or overestimating 5-HT_{1A} or 5-HT_{1B} receptor selectivity was observed for a series of serotonin receptor agonists. Antagonist receptor selectivity was not affected significantly by the nature of the 5-HT₂ receptor assay used. These data indicate that, by using an antagonist radioligand to label 5-HT₂ receptors and agonist radioligands to label 5-HT₁ receptors, the 5-HT₁ receptor selectivity may be overestimated. This may be an especially severe problem in serotonin drug development as drugs that interact potently with 5-HT₂ receptors have been reported to be psychoactive and/or hallucinogenic.

Evidence for the existence of multiple functional serotonin receptors in the brain and periphery has been accumulating rapidly [1, 2]. 5-HT₂ receptors have been labeled with [³H]spiperone [3] and [³H]ketanserin [4, 5] which are antagonist radioligands. [³H]Serotonin-radiolabeling studies have revealed three distinct sites, each possessing characteristics indicating that they may be functional receptors [6-8]. To date, the majority of studies have used the agonist radioligand [³H]-8-OH-DPAT§ to label 5-HT_{1A} sites [6], [³H]-5-HT to label the 5-HT_{1A}, 5-HT_{1B}, and 5-HT_{1C} sites and [³H]mesulergine to label 5-HT_{1C} sites [2, 8-10]. In studies aimed at assessing the relative affinities of serotonergic compounds for 5-HT₂, 5-HT_{1A}, 5-HT_{1B}, and 5-HT_{1C} receptors, laboratories have been using antagonist radioligands to label the 5-HT₂ receptors and agonist radioligands to label the multiple 5-HT₁ receptors [8-10].

Studies from our laboratory have demonstrated that the 5-HT₂ receptor apparently exists in two states, both labeled with equal affinity by the antagonist radioligand [³H]ketanserin [5, 11, 12]. Apparent agonist affinities for the receptor appear to be a combination of their affinities for the two states of the receptor for which the agonists have very different affinities. The resultant apparent affinity is predominantly influenced by the low agonist affinity state of the receptor as this state appears to exist in excess of the high agonist affinity state of the receptor [5, 11]. Assays labeling 5-HT_{1A}, 5-HT_{1B}, and 5-HT_{1C} receptors do not involve the low affinity state of these receptors as agonist radioligands selectively label the agonist high affinity state of the receptor as revealed by the high affinity serotonin displays in radioligand binding assays of these sites. Therefore, an agonist radioligand that would selectively label the agonist high affinity state of the 5-HT₂ receptor would be a useful tool in determining true relative affinities of drugs for the multiple serotonin receptors (i.e. 5-HT_{1A} vs 5-HT₂). [³H]DOB, a putative potent and specific 5-HT₂ agonist radioligand, has been developed [13-15]. We report herein results of studies using [³H]DOB, in conjunction with other radio-

† To whom all correspondence should be sent.

§ Abbreviations: 8-OH-DPAT, 8-hydroxy-2-(di-*n*-propylamino)-tetralin; DOB, 4-bromo-2,5-dimethoxyphenylisopropylamine; TFMPP, 1-(3-trifluoromethylphenyl)piperazine; and GppNHp, guanylyl-5'-imidodiphosphate.

ligand assays, to screen serotonergic drugs for their selectivities for agonist radiolabeled 5-HT₂, 5-HT_{1A}, 5-HT_{1B}, and 5-HT_{1C} receptors.

MATERIALS AND METHODS

Tissue preparation. Following decapitation, the brains of male Sprague-Dawley rats were removed and placed in 0.9% ice-cold saline and then dissected over ice until the tissue was prepared. Tissues were stored in ice-cold saline for no longer than 1 hr and, following blot drying and weighing, they were prepared and frozen at -30° until used. Freshly dissected tissue was homogenized (polytron setting 6 for 20 sec) in 30 vol. of ice-cold buffer containing 50 mM Tris-HCl (pH 7.4 at 37° ; pH 8.0 at 4°), 0.5 mM Na₂EDTA and 10 mM MgSO₄ and centrifuged at 30,000 *g* for 15 min. The supernatant fraction was discarded, and the pellet was resuspended and pre-incubated for 15 min at 37° . The homogenate membranes were washed twice by centrifugation and resuspension. The final suspension buffer contained 10 μ M pargyline and 0.1% ascorbate and was added last to the assay incubation medium. Protein determinations were made by the method of Lowry *et al.* [16].

Radioligand binding assays. The agonist high affinity state of the 5-HT₂ receptor was labeled with 0.4 nM [³H]DOB (40 Ci/mmol, New England Nuclear) using 20 mg of membrane suspension prepared from rat frontal cortical tissue [13,14]. Cinanserin (10^{-6} M) was used to define non-specific binding. The agonist low affinity state of the 5-HT₂ receptor was assayed with 0.4 nM [³H]ketanserin (76 Ci/mmol, New England Nuclear) and 3 mg of homogenized rat frontal cortical tissue in the presence of 10^{-5} M GppNHp [5]. In addition, the results of agonist competition for [³H]ketanserin-labeled 5-HT₂ receptors in the absence of GppNHp were

computer-analyzed (see below) to determine the agonist affinity for the agonist low affinity state. The 5-HT_{1A} receptor was labeled with 0.1 nM [³H]-8-OH-DPAT (120 Ci/mmol, New England Nuclear) using 6 mg of rat hippocampal homogenates. 8-OH-DPAT (10^{-6} M) was used to define nonspecific binding. The 5-HT_{1B} receptor was labeled with 2.0 nM [³H]serotonin (23 Ci/mmol, New England Nuclear) using 8 mg of rat striatal membrane homogenates. 5-HT (10^{-6} M) was used to define non-specific binding, and 10^{-7} M 8-OH-DPAT and mesulergine [6, 8] were included to block 5-HT_{1A} and 5-HT_{1C} receptors respectively. The 5-HT_{1C} receptor was labeled with 1.0 nM [³H]mesulergine (85 Ci/mmol, Amersham) using 20 mg of pig cortical homogenate in the presence of 20 nM spiperone. Non-specific binding was determined with 10 μ M serotonin. Eleven concentrations of non-radioactive competing drugs were made fresh daily in assay buffer. Following incubation with membranes and radioligand at 37° for 15 min (for 5-HT₂ assays) or 30 min (for 5-HT₁ assays), samples were rapidly filtered over glass fiber filters and washed with 10 ml of ice-cold 50 mM Tris-HCl buffer. Individual filters were inserted into vials and equilibrated with 5 ml of scintillation fluid for 6 hr before counting at 50% efficiency. Results were analyzed using an updated version of the program EBDA [17] or RS1 (BBN software). EBDA is a program for the IBM PC that uses non-linear regression to best-fit the competition curve data to the Hill equation [17]. RS1 is a graphics program for the IBM AT that was used to plot the competition curves according to the parameters determined using EBDA.

RESULTS

The results presented in Figs. 1–3 demonstrate that the apparent affinity of serotonin and two putative

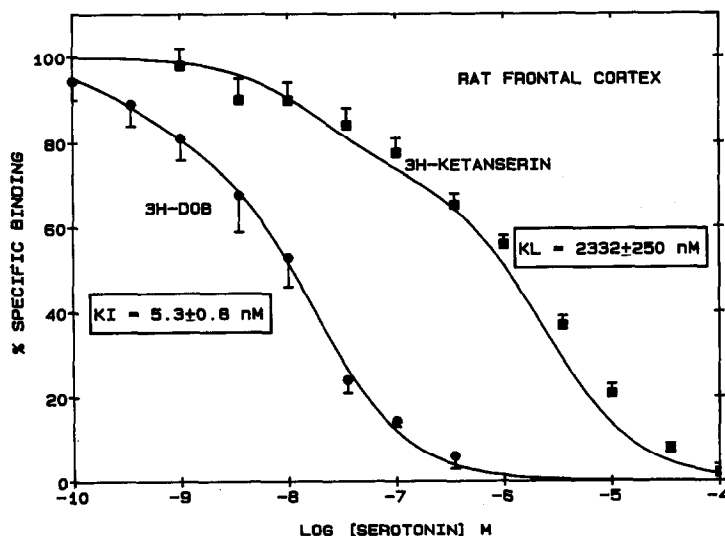


Fig. 1. Serotonin competition for 5-HT₂ receptors labeled with 0.4 nM [³H]DOB or 0.4 nM [³H]ketanserin. Total dpm for [³H]DOB was 1180 ± 37 of which 54% was specific. Total dpm for [³H]ketanserin was 4100 ± 380 of which 84% was specific. The K_L values refer to agonist affinities for the low agonist affinity state of the [³H]ketanserin-labeled 5-HT₂ receptor and were determined using computer-assisted analysis of a two-site model (see Material and Methods). Error bars are the SEM from three independent experiments, each point determined in triplicate.

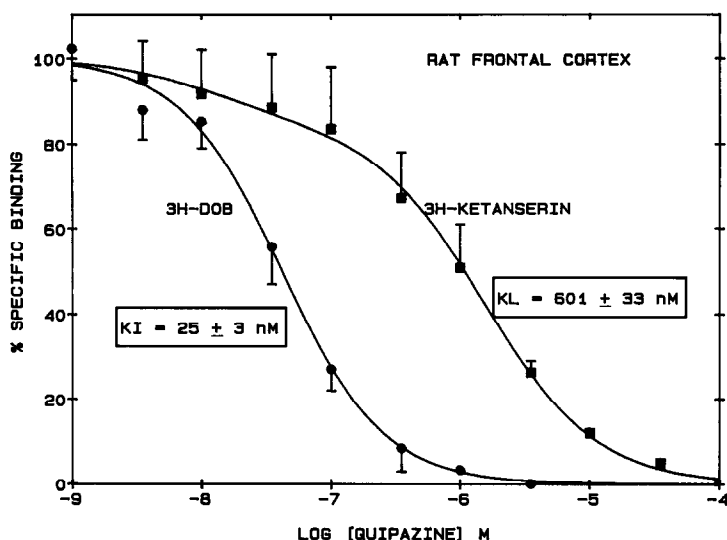


Fig. 2. Quipazine competition for 5-HT₂ receptors labeled with 0.4 nM [³H]DOB or 0.4 nM [³H]ketanserin (see legend of Fig. 1). K_L was determined by computer-assisted analysis of a two-site model.

serotonin receptor agonists, quipazine and TFMPP, for the radiolabeled 5-HT₂ receptor is dependent on the agonist/antagonist nature of the radioligand used. In assays using [³H]DOB, an agonist radioligand for 5-HT₂ receptors [13, 14], serotonin, quipazine, and TFMPP produced potent competition curves, far to the left of the competition curve produced by these agonists in competing for the 5-HT₂ antagonist radioligand [³H]ketanserin. The high affinities of serotonin and DOB for the [³H]DOB-labeled 5-HT₂ receptor were similar to the high affinities of these substances previously reported using computer-assisted two-site analysis of [³H]ketanserin binding [5, 12]. However, the competition curves produced by the antagonists ketan-

serin and propranolol appeared to be independent of the radioligand used to label the 5-HT₂ receptor (Figs. 4 and 5). Chlorpromazine demonstrated a marginally higher affinity (4- to 5-fold) for the [³H]ketanserin-labeled 5-HT₂ receptor than for the [³H]DOB-labeled 5-HT₂ receptor. The exact meaning of this anomalous observation is not clear.

The K_I values determined for a series of agonists at the two states of the 5-HT₂ receptor and three 5-HT₁ receptor sub-types are listed in Table 1. Table 2 lists the affinity ratios for the two states of the 5-HT₂ receptor and illustrates that, using the K_I values determined with the 5-HT₂ antagonist radioligand, the potency ratios of agonists for the 5-HT_{1A}, 5-HT_{1B}, and 5-HT_{1C} receptor sub-types were dra-

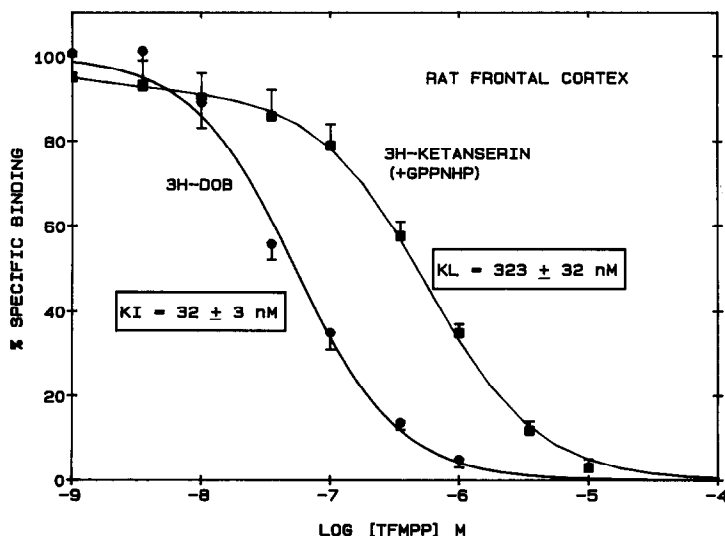


Fig. 3. TFMPP competition for 5-HT₂ receptors labeled with 0.4 nM [³H]DOB or 0.4 nM [³H]ketanserin in the presence of 10⁻⁵ M GppNHp (see legend of Fig. 1). The [³H]ketanserin K_L value represents the affinity of TFMPP for the agonist low affinity state of the 5-HT₂ receptor (see Materials and Methods).

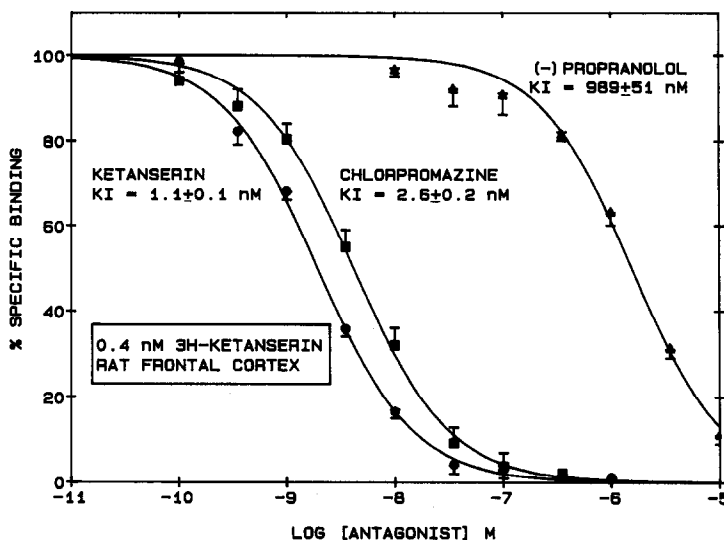


Fig. 4. Antagonist competition for 5-HT₂ receptors labeled with 0.4 nM [³H]ketanserin (see legend of Fig. 1).

matically different than the potency ratios calculated using the K_i values determined using the 5-HT₂ agonist radioligand. Antagonist potency ratios appear to be relatively independent of the nature of the 5-HT₂ radioligand used.

DISCUSSION

There is a great deal of interest in developing drugs that are specific agonists at various serotonin receptors, using radioligand binding assays as screening tools [9, 18]. The data displayed herein indicate that the apparent affinity of putative serotonergic agents for the 5-HT₂ receptor was dependent on the nature (intrinsic activity) of the radioligand. Agonists appeared to have 50- to 100-fold higher affinity when

the agonist radioligand [³H]DOB was used to label the receptor than when the antagonist radioligand [³H]ketanserin was used. This observation is strongly supportive of, and consistent with, previous work from our laboratory, indicating the presence of agonist high and low affinity states for the 5-HT₂ receptor due to 5-HT₂ receptor/GTP-binding protein interactions [5, 11, 12, 14]. The 5-HT_{1A}, 5-HT_{1B}, and 5-HT_{1C} receptor sub-types are typically labeled with radioactive ligands that appear to label the agonist high affinity states of these receptors. As a result, when affinities of putative agonists are compared at ³H-antagonist-labeled 5-HT₂ receptors and ³H-agonist-labeled 5-HT₁ receptor sub-types, the 5-HT₁ selectivity may be greatly overestimated.

Recent evidence has indicated the distinct possi-

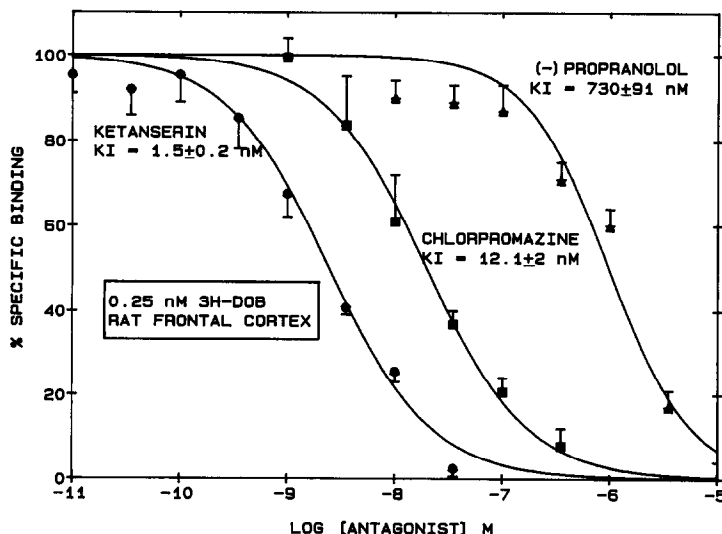


Fig. 5. Antagonist competition for 5-HT₂ receptors labeled with 0.4 nM [³H]DOB (see legend of Fig. 1).

Table 1. Affinities of selected agonists and antagonists for radiolabeled 5-HT₂ and 5-HT₁ receptors

	<i>K_i</i> (nM)				
	5-HT _{2H}	5-HT _{2L}	5-HT _{1A}	5-HT _{1B}	5-HT _{1C}
Agonists					
Serotonin	5.3 ± 0.8	2,332 ± 250	1.7 ± 0.3	5.4 ± 0.5	9.5 ± 1.5
Quipazine	25 ± 2	601 ± 33	2,038 ± 84	191 ± 36	624 ± 87
TFMPP	32 ± 3	323 ± 32	173 ± 12	27 ± 4	120 ± 12
(±)DOB	0.44 ± 0.05	59 ± 4	3,770 ± 118	831 ± 37	69 ± 16
Buspirone	289 ± 38	576 ± 20	14 ± 1.0	>10,000	1,160 ± 81
Ipsapirone	1,047 ± 55	1,305 ± 31	5.5 ± 0.3	>10,000	3,306 ± 325
RU 24969	41 ± 5	1,860 ± 464	3.8 ± 0.3	0.91 ± 0.15	560 ± 70
8-OH-DPAT	696 ± 112	5,350 ± 374	1.2 ± 0.12	>10,000	5,500 ± 100
Antagonists					
Spiperone	0.8 ± 0.04	0.42 ± 0.04	101 ± 9	>10,000	62 ± 6
Ketanserin	1.5 ± 0.2	1.1 ± 0.1	1,933 ± 219	>10,000	49 ± 4
Chlorpromazine	12.1 ± 2	2.6 ± 0.2	737 ± 72	1,489 ± 101	30 ± 3
(-)-Propanolol	730 ± 91	989 ± 51	55 ± 4	17 ± 4	1,292 ± 78
(±)Pindolol	4,542 ± 570	16,851 ± 2,600	31 ± 4	34 ± 3	>10,000

The agonist high affinity state (5-HT_{2H}) of the 5-HT₂ receptor was labeled with the agonist radioligand [³H]DOB; the agonist low affinity state (5-HT_{2L}) with [³H]ketanserin, the 5-HT_{1A} receptor with [³H]-8-OH-DPAT, the 5-HT_{1B} receptor with [³H]-5-HT, and the 5-HT_{1C} receptor with [³H]mesulergine. Values are the means ± SEM of three independent experiments.

bility that brain 5-HT₂ receptor activation may be the causative factor in the hallucinogenic (psychoactive) effects of many drugs of abuse, including LSD [12, 19]. Therefore, it seems especially important in serotonin receptor drug development to not underestimate the 5-HT₂ receptor affinity of novel compounds. The agonist radioligand [³H]DOB appears to be a screening tool that will avoid this complication.

One important implication of the data presented in Tables 1 and 2 is that the early pharmacological definition of 5-HT₂ receptors as possessing micro-

molar affinity for serotonin was an over-simplification [1]. Serotonin displays a *K_i* value of 5.3 nM for [³H]DOB-labeled 5-HT₂ receptors (Table 1). A question raised by these results is why [³H]serotonin does not label 5-HT₂ receptors. Studies in our laboratory (M. Titeler and G. Battaglia, unpublished observations) demonstrated that approximately 5–10% of specific [³H]serotonin binding was potentially inhibited by the 5-HT₂ receptor antagonists spiperone, cinanserin, and ketanserin. The apparent reason for such a low level of 5-HT₂ receptor binding is that, relative to the total amounts

Table 2. 5-HT₂:5-HT₁ serotonin receptor selectivities of serotonergic drugs

	Ratio of K_i values						
	5-HT _{2L}	5-HT _{2H}	5-HT _{2L}	5-HT _{2H}	5-HT _{2L}	5-HT _{2H}	5-HT _{2L}
	5-HT _{2H}	5-HT _{1A}	5-HT _{1A}	5-HT _{1B}	5-HT _{1B}	5-HT _{1C}	5-HT _{1C}
Agonists							
Serotonin	440	3.1	1372	0.98	432	0.56	245
Quipazine	24	0.012	0.29	0.13	3.1	0.04	0.96
TFMPP	10	0.18	1.87	1.19	12	0.27	2.69
(±)DOB	134	1.2×10^{-4}	0.016	5.3×10^{-4}	0.07	0.006	0.86
Buspirone	2.0	21	41	21	0.058	0.25	0.50
Ipsapirone	1.2	190	237	190	0.13	0.32	0.39
RU 24969	45	10.8	489	10.8	2044	0.07	3.32
8-OH-DPAT	7.7	580	4458	580	0.54	0.13	0.97
Antagonists							
Spiperone	0.5	7.9×10^{-3}	4.2×10^{-3}	8×10^{-5}	4.2×10^{-5}	0.13	0.01
Ketanserin	0.7	7.8×10^{-4}	5.7×10^{-4}	1.1×10^{-4}	1.1×10^{-4}	0.03	0.02
Chlorpromazine	0.2	9.5×10^{-3}	3.5×10^{-3}	4.7×10^{-3}	1.7×10^{-3}	0.40	0.09
(-)-Propranolol	1.4	13.3	18	43	58	0.59	0.77
(±)Pindolol	3.7	147	544	134	496	0.45	1.69

The selectivity was quantified by determining the ratio of affinities (*K_i* values) of drugs for the two states of the 5-HT₂ receptor and 5-HT_{1A}, 5-HT_{1B}, and 5-HT_{1C} receptors (from Table 1). Note the influence of [³H]DOB versus [³H]ketanserin on the apparent selectivity. In column 1, the higher the ratio the more selective for the 5-HT_{2H} versus the 5-HT_{2L} state the drug appears to be. In columns 2–7, the larger the ratio the more selective for the 5-HT_{1A}, 5-HT_{1B}, or 5-HT_{1C} receptor the drug appears to be. The smaller the ratio the more selective for the 5-HT₂ receptor the drug appears to be.

of available receptor sites with high affinity for serotonin, the amount of agonist high affinity state of the 5-HT₂ receptor is very low [13, 14].

The results with quipazine help to explain the observed selectivity of this drug as a 5-HT₂ agonist *in vivo*. When [³H]ketanserin was used as the radioligand, quipazine appeared to have higher affinity for 5-HT_{1B} sites than for 5-HT₂ and similar affinities for 5-HT_{1A} and 5-HT₂ receptors. However, quipazine has been found to be a potent and specific 5-HT₂ receptor agonist *in vivo* [18, 20]. When [³H]DOB was used as the 5-HT₂ radioligand, quipazine appeared to be 10- to 100-fold less potent at the 5-HT_{1A} and 5-HT_{1B} sites than at the 5-HT₂ receptor. Thus, the use of [³H]DOB as the radioligand apparently resulted in the determination of an affinity constant for quipazine which more accurately predicts its receptor specificity *in vivo*. Several interesting speculations may be made as a result of the data in Tables 1 and 2. TFMPP is a putative specific 5-HT_{1B} agonist [9]. However, when [³H]DOB was used as the radioligand, the affinities of TFMPP for the 5-HT_{1B} and 5-HT₂ receptors were essentially identical. It would be reasonable to predict that the biological actions of TFMPP may be a result of stimulation of both 5-HT_{1B} and 5-HT₂ receptors, if indeed TFMPP is an agonist at the 5-HT₂ receptor (see below). Buspirone and ipsapirone (formerly TVX Q 7821) are putative 5-HT_{1A} agonists with anxiolytic effects [21, 22]. The data in Table 1 indicate that at 5-HT₂ receptors these drugs have similar apparent affinities whether [³H]DOB or [³H]ketanserin is used as the radioligand. This may indicate that these drugs are antagonists at the 5-HT₂ receptor as this pattern of binding is observed with well-characterized 5-HT₂ antagonists (Table 1). However, it is unlikely that the effects of buspirone or ipsapirone *in vivo* involve 5-HT₂ interactions as the potencies of these drugs at the 5-HT₂ receptor were much lower than their potencies at the 5-HT_{1A} receptor (Table 2).

It should be noted that the division of drugs into agonists and antagonists (Tables 1 and 2) is based on various criteria. For instance, quipazine [20], TFMPP [23], DOB [24], 8-OH-DPAT [25], and RU 24969 [26] have been shown to produce behavioural responses and/or neurochemical effects on brain serotonergic systems consistent with the direct stimulation of some class or classes of brain serotonin receptors. Buspirone [27] and ipsapirone [28] have been shown to potentially inhibit firing of brain serotonergic neurons through stimulation of 5-HT_{1A} receptors on cell bodies in the dorsal raphe nucleus. The fact that a drug stimulates one serotonin receptor does not preclude the possibility that it may act as an antagonist at another serotonin receptor. For instance, while TFMPP clearly produces serotonin receptor agonist mediated effects, presumably through 5-HT_{1B} receptors [9], there is evidence that it may act as an antagonist at 5-HT₂ receptors [29]. As mentioned above, the data presented herein indicate the possibility that ipsapirone and buspirone may act as 5-HT₂ receptor antagonists as well as 5-HT_{1A} agonists. The classification of spiperone, ketanserin, and chlorpromazine as antagonists is based on reports demonstrating the blockade of well-

studied serotonergic behaviors in rats [3]. Propranolol and pindolol are classical beta-adrenergic receptor antagonists [30] that have been shown to potentially interact with brain serotonin systems [31–33]. These studies indicate that propranolol and/or pindolol have antagonist properties at 5-HT_{1A} receptors although some agonist-like effects have been noted. The classification scheme used in Tables 1 and 2 is meant to indicate the major pharmacological effect of the drug reported by serotonin receptor pharmacologists.

Acknowledgement—This work was supported by PHS Grant MH40716-01A2.

REFERENCES

1. S. J. Peroutka, R. M. Lebovitz and S. H. Snyder, *Science* **212**, 827 (1981).
2. N. W. Pedigo, H. I. Yamamura and D. L. Nelson, *J. Neurochem.* **36**, 220 (1981).
3. J. E. Leysen, C. J. E. Niemegeers, J. P. Tollenaere and P. M. Laduron, *Nature, Lond.* **272**, 168 (1978).
4. J. E. Leysen, C. J. E. Niemegeers, J. M. Van Nueten and P. M. Laduron, *Molec. Pharmac.* **21**, 301 (1982).
5. G. Battaglia, M. Shannon, N. Borgundvaag and M. Titeler, *J. Neurochem.* **43**, 1213 (1984).
6. M. D. Hall, S. El Mestikawy, M. B. Emerit, L. Pichat, M. Hamon and H. Gozlan, *J. Neurochem.* **44**, 1685 (1985).
7. K. A. Yagaloff and P. R. Hartig, *J. Neurosci.* **5**, 3178 (1985).
8. D. Hoyer, S. Srivatsa, A. Pazos, G. Engel and J. M. Palacios, *Neurosci. Lett.* **69**, 269 (1986).
9. M. A. Sills, B. B. Wolfe and A. Frazer, *J. Pharmac. exp. Ther.* **231**, 480 (1984).
10. S. J. Peroutka, *J. Neurochem.* **47**, 529 (1986).
11. M. Titeler, G. Battaglia and M. Shannon, *J. Receptor Res.* **7**, 705 (1984).
12. M. Shannon, G. Battaglia, R. A. Glennon and M. Titeler, *Eur. J. Pharmac.* **102**, 23 (1984).
13. M. Titeler, K. Herrick, R. A. Lyon, J. D. McKenney and R. A. Glennon, *Eur. J. Pharmac.* **117**, 145 (1985).
14. R. A. Lyon, K. Davis and M. Titeler, *Molec. Pharmac.* **31**, 194 (1987).
15. R. A. Glennon, J. D. McKenney, R. A. Lyon and M. Titeler, *J. med. Chem.* **29**, 914 (1986).
16. O. H. Lowry, N. J. Rosebrough, A. L. Farr and R. J. Randall, *J. biol. Chem.* **193**, 265 (1951).
17. G. A. Macpherson, *Comput. Prog. Biomed.* **17**, 107 (1983).
18. R. A. Glennon, R. M. Slusher, R. A. Lyon, M. Titeler and J. D. McKenney, *J. med. Chem.* **29**, 2375 (1986).
19. R. A. Glennon and M. Titeler, *Life Sci.* **35**, 2505 (1984).
20. R. L. Friedman, R. J. Barrett and E. J. Sanders-Bush, *J. Pharmac. exp. Ther.* **228**, 628 (1984).
21. L. A. Riblet, A. S. Eison, M. S. Eison, D. P. Taylor, D. L. Temple and C. P. VanderMaalen, *Psychopathology*, **17**(Suppl. 3), 69 (1984).
22. W. U. Dompert, T. Glaser and J. Traber, *Naunyn-Schmiedeberg's Archs Pharmac.* **328**, 467 (1985).
23. L. S. Brady and J. E. Barrett, *J. Pharmac. exp. Ther.* **235**, 436 (1985).
24. R. A. Glennon, R. Young, J. M. Jacyno, M. Slusher and J. A. Rosecran, *Eur. J. Pharmac.* **86**, 453 (1983).
25. R. J. De Souza, G. M. Goodwin, A. R. Green and D. J. Heal, *Br. J. Pharmac.* **89**, 377 (1986).
26. C. Euvrard and J. R. Biossier, *Eur. J. Pharmac.* **63**, 65 (1980).

27. C. P. VanderMaelen, G. K. Matheson, R. C. Wilderman and L. A. Patterson, *Eur. J. Pharmac.* **129**, 123 (1986).
28. A. Basse-Tomsuk and G. V. Rebec, *Eur. J. Pharmac.* **130**, 141 (1986).
29. E. Sanders-Bush and P. J. Conn, *Psychopharmac. Bull.* **22**, 829 (1986).
30. K. P. Minneman, L. R. Hegstrand and P. B. Molinoff, *Molec. Pharmac.* **16**, 34 (1979).
31. S. Hjorth and A. Carlsson, *Neuropharmacology* **24**, 1143 (1985).
32. J. S. Sprouse and G. K. Aghajanian, *Eur. J. Pharmac.* **128**, 295 (1986).
33. S. Hjorth and A. Carlsson, *Eur. J. Pharmac.* **129**, 131 (1986).